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=> isotype matched negative control
L1 0 FILE AGRICOLA
L2 1 FILE BIOTECHNO
L3 0 FILE CONFSCI
L4 0 FILE HEALSAFE
L5 0 FILE IMSDRUGCONF
L6 1 FILE LIFESCI
L7 0 FILE MEDICONF
L8 2 FILE PASCAL

TOTAL FOR ALL FILES
L9 4 ISOTYPE MATCHED NEGATIVE CONTROL

=> dup rem
ENTER L# LIST OR (END):l9
DUPLICATE IS NOT AVAILABLE IN 'IMSDRUGCONF, MEDICONF'.
ANSWERS FROM THESE FILES WILL BE CONSIDERED UNIQUE
PROCESSING COMPLETED FOR L9
L10 2 DUP REM L9 (2 DUPLICATES REMOVED)

=> d l10 ibib abs total

L10 ANSWER 1 OF 2 PASCAL COPYRIGHT 2003 INIST-CNRS. ALL RIGHTS RESERVED. on STN
ACCESSION NUMBER: 2002-0555334 PASCAL
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reserved.
TITLE (IN ENGLISH): GLUT1 antibody staining in thin-layer specimens of benign and malignant body cavity effusions
AUTHOR: MENSCH Leon S.; WELLER Lindsay; SIMMONS-ARNOLD Linda; GIBSON Pamela C.; LEIMAN Gladwyn; BEATTY Barbara
CORPORATE SOURCE: Divisions of Experimental Pathology and Cytopathology, Department of Anatomical Pathology, Fletcher Allen Health Care and University of Vermont, Burlington, Vermont, United States
SOURCE: Acta cytologica, (2002), 46(5), 813-818, 9 refs.
ISSN: 0001-5547 CODEN: ACYTAN
DOCUMENT TYPE: Journal
BIBLIOGRAPHIC LEVEL: Analytic
COUNTRY: United States
LANGUAGE: English
AVAILABILITY: INIST-10515, 354000109305770040
AN 2002-0555334 PASCAL
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AB OBJECTIVE: To determine whether GLUT1 antibody could replace one or more of the currently used antiepithelial antibodies and to assess whether ThinPrep.RTM. methodology is suited to immunocytochemical (ICC) evaluation. STUDY DESIGN: In a prospective study of 10 fluids containing malignant cells from cases of proven adenocarcinoma and 10 cytologically benign effusions, multiple slides were prepared by ThinPrep.RTM. technology for staining with four commercially available antibodies and appropriate **isotype-matched negative controls**. The antibodies used were GLUT1, CEA, B72.3 and Leu-M1 (CD 15). Tissue sections and ThinPrep.RTM. slides were used as positive controls. Specimens were batched to ensure similar conditions for all antibody reactions. RESULTS: Of the 11 cases ultimately proven to be carcinoma, GLUT1 and B72.3 stained 7 each (63.6%), and CEA and Leu-M1 6 each (54.5%). No false positive staining was encountered, but one case chosen as a benign control was shown to contain immunopositive cells by three of the four epithelial markers used; this case was therefore an occult true positive rather than a false positive. CONCLUSION: In this small but controlled prospective analysis, GLUT1 demonstrated strong positive staining, with sensitivity similar to that of currently used epithelial markers. Using GLUT1 in conjunction with B72.3, no cases of carcinoma were missed. GLUT1 could be used in a panel of antibodies designed to confirm the presence of adenocarcinoma. ThinPrep.RTM. methodology, which enables multiple slides to be prepared after routine microscopy determines the need for ICC, appears suited to this adjuvant investigation.

L10 ANSWER 2 OF 2 BIOTECHNO COPYRIGHT 2003 Elsevier Science B.V. on STN
DUPLICATE
ACCESSION NUMBER: 2000:30734596 BIOTECHNO
TITLE: Intracellular markers
AUTHOR: Koester S.K.; Bolton W.E.
CORPORATE SOURCE: W.E. Bolton, Beckman Coulter, Inc., Advanced Technology, M/C 22-A01, 11800 S.W. 147th Avenue, Miami, FL 33196-2500, United States.
E-mail: wadeb4@aol.com
SOURCE: Journal of Immunological Methods, (21 SEP 2000), 243/1-2 (99-106), 54 reference(s)
CODEN: JIMMBG ISSN: 0022-1759
PUBLISHER ITEM IDENT.: S0022175900002398
DOCUMENT TYPE: Journal; General Review
COUNTRY: Netherlands
LANGUAGE: English
SUMMARY LANGUAGE: English
AN 2000:30734596 BIOTECHNO
AB An increased level of complexity will be encountered when developing protocols for intracellular markers. Protocols for surface markers have

been successfully standardized, however it is understood that no single method is appropriate for all intracellular staining. A systematic approach should be followed, including knowledge of antigen location and functional state, selection of cell fixative and cell permeabilizer, antibody specificity and class/subclass, fluorochrome, fluorochrome to protein ratio (F:P), and use of adequate controls, including **isotype-matched negative controls** and positive and negative cell controls. Even though it is impossible to recommend a single technique to stain all intracellular antigens, the authors present a logical approach to follow when developing a staining protocol. (C) 2000 Elsevier Science B.V.

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FULL ESTIMATED COST                                       9.13       9.34

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=> isotype matched negative control
L11           2 FILE CAPLUS
L12           1 FILE BIOTECHNO
L13           0 FILE COMPENDEX
L14           0 FILE ANABSTR
L15           0 FILE CERAB
L16           0 FILE METADEX
L17          91 FILE USPATFULL

TOTAL FOR ALL FILES
L18          94 ISOTYPE MATCHED NEGATIVE CONTROL

=> dup rem
ENTER L# LIST OR (END):l11-16
L13 HAS NO ANSWERS
L14 HAS NO ANSWERS
L15 HAS NO ANSWERS
L16 HAS NO ANSWERS
PROCESSING COMPLETED FOR L11
PROCESSING COMPLETED FOR L12
PROCESSING COMPLETED FOR L13
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PROCESSING COMPLETED FOR L14
PROCESSING COMPLETED FOR L15
PROCESSING COMPLETED FOR L16
L19 2 DUP REM L11-16 (1 DUPLICATE REMOVED)

=> d l19 ibib abs total

L19 ANSWER 1 OF 2 CAPLUS COPYRIGHT 2003 ACS on STN DUPLICATE 1
ACCESSION NUMBER: 2000:635692 CAPLUS
DOCUMENT NUMBER: 134:28283
TITLE: Intracellular markers
AUTHOR(S): Koester, S. K.; Bolton, W. E.
CORPORATE SOURCE: Advanced Technology, Beckman Coulter, Inc., Miami, FL, 33196-2500, USA
SOURCE: Journal of Immunological Methods (2000), 243(1-2), 99-106
CODEN: JIMMBG; ISSN: 0022-1759
PUBLISHER: Elsevier Science B.V.
DOCUMENT TYPE: Journal; General Review
LANGUAGE: English
AB A review with numerous refs. An increased level of complexity will be encountered when developing protocols for intracellular markers. Protocols for surface markers have been successfully standardized, however it is understood that no single method is appropriate for all intracellular staining. A systematic approach should be followed, including knowledge of antigen location and functional state, selection of cell fixative and cell permeabilizer, antibody specificity and class/subclass, fluorochrome, fluorochrome to protein ratio (F:P), and use of adequate controls, including **isotype-matched neg. controls** and pos. and neg. cell controls. Even though it is impossible to recommend a single technique to stain all intracellular antigens, the authors present a logical approach to follow when developing a staining protocol.
REFERENCE COUNT: 54 THERE ARE 54 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L19 ANSWER 2 OF 2 CAPLUS COPYRIGHT 2003 ACS on STN
ACCESSION NUMBER: 1993:93649 CAPLUS
DOCUMENT NUMBER: 118:93649
TITLE: In vitro tumor toxicity with antibody-toxin conjugates
AUTHOR(S): May, Richard D.; Fulton, R. Jerrold
CORPORATE SOURCE: Chemotherapy Toxicol. Dep., South. Res. Inst., Birmingham, AL, USA
SOURCE: In Vitro Methods Toxicol. (1992), 9-20. Editor(s): Watson, Ronald Ross. CRC: Boca Raton, Fla.
CODEN: 58SAA7
DOCUMENT TYPE: Conference; General Review
LANGUAGE: English
AB A review with 79 refs. in how to prep. and test immunotoxins (ITs) for killing tumor cells in vitro. The section of toxin and Ab are crucial for constructing a successful IT. The authors have provided detailed methodol. for constructing an IT-A using ricin A chain, which is widely used in this field. The purity, Ab binding activity, and cytotoxic efficacy of the IT-As are tested by appropriate in vitro methods. Specificity of killing is demonstrated by a direct IT-A assay, which includes using an **isotype-matched neg. control** IT-A, as well as an Ag-neg. cell line. Careful in vitro testing of ITs can predict some of the problems assocd. with their in vivo applications, such as liver toxicity, and nonspecific binding to nontarget tissues. However, in vitro efficacy may not always be translated into in vivo usefulness.'thus, preclin. in vivo animal models must eventually be developed and tested prior to using ITs clin.